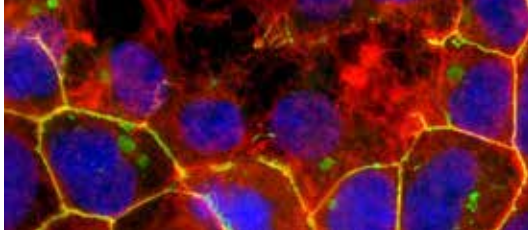
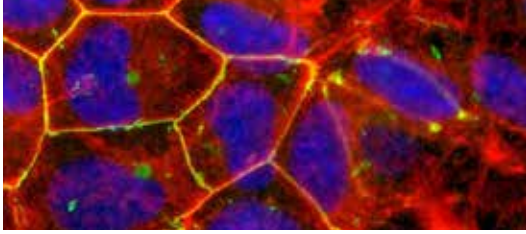
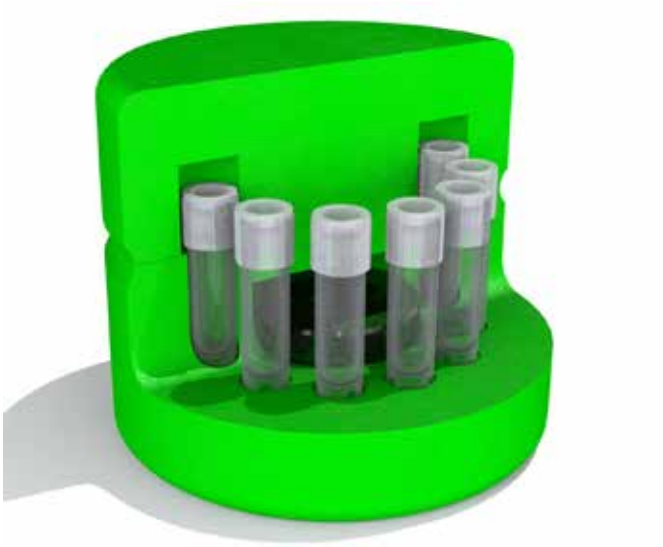
THE ESSENTIALS OF
LIFE SCIENCE RESEARCH
GLOBALLY DELIVERED™





COOLCELL® LX ALCOHOL-FREE CRYOPRESERVATION CONTAINERS ATCC® No. ACS-6000

CoolCell® LX cryopreservation containers reliably provide the ideal conditions (-1˚C per minute freezing rate when placed at -80˚C) for the safe and effective cryopreservation of your cells. Alcohol is not required; simply fill the CoolCell LX container, with your cryovials, and place it in a -80˚C freezer. The new beveled design of the CoolCell LX lid, allows for easy opening and access to the cryovials, simplifying their transfer to long-term storage. At the same time, the numbered vial holes allow for quick indexing of cryovials, helping you to keep your samples organized. CoolCell LX container accommodates up to 12 cryovials (1 to 2 mL cryovials).



OTHER FEATURES OF THE COOLCELL LX CONTAINER INCLUDE:

- High post-thaw viability and performance
- Reproducible cryopreservation conditions
- No on-going cost and no maintenance

The solid-state core and highly insulative outer material precisely balance heat removal during the freezing period to ensure repeatable, consistent cooling all the way to the final cryogenic storage temperature.

COOLCELL LX CRYOPRESERVATION CONTAINER IS PROVEN FOR USE WITH A VARIETY OF CELLS INCLUDING:

Stem Cell/ Pre-Stem Cell	Primary Cells	Continuous Cell Lines
Preadipocytes	Neonatal Keratinocytes	CHO
Breast Cancer Stem Cells	Human and Mouse White Blood Cells	LnCap
Colon Cancer Stem Cells	Human CD34+ Cells	HTB77
Glioblastoma Cancer Stem Cells	Muscle Cells	A549
Mouse Embryonic Stem Cells	Human Tendon Fibroblasts	HeLa
Human Endothelial Progenitor Cells	Melanoma Tumor Cells	
	Human Ventricular and Atrial Cardiac Cells	

PRIMARY CELLS

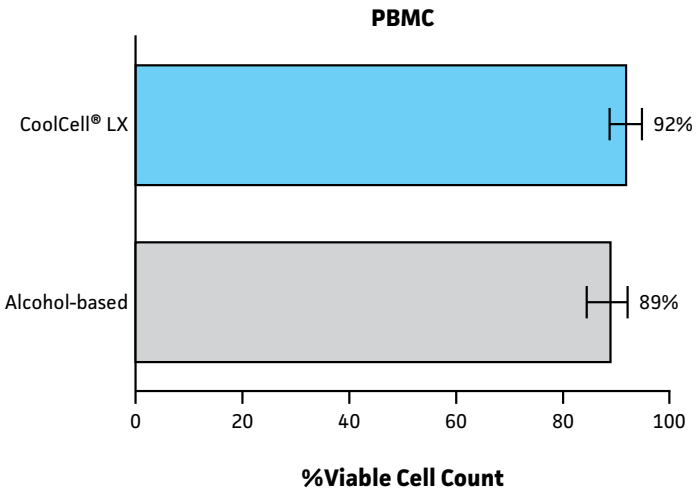


Figure 1. Post-thaw samples of PBMC cells, cryopreserved using either the CoolCell® LX or the Mr. Frosty® isopropanol-containing chamber, were tested for viability using the trypan blue method. CoolCell LX performed as well as Mr. Frosty, without the requirement for alcohol.

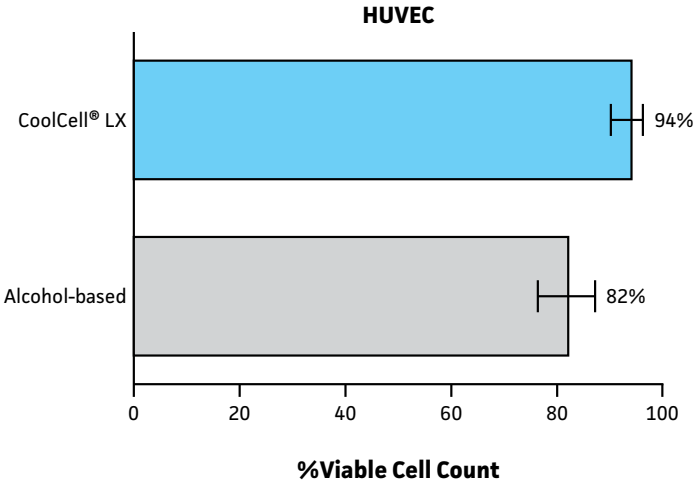


Figure 2. Post-thaw samples of HUVECs, cryopreserved using either the CoolCell® LX or the Mr. Frosty® isopropanol-containing chamber were tested for viability using the trypan blue method. Post-thaw viability of HUVECs was markedly higher in samples cryopreserved using the CoolCell LX compared to samples cryopreserved using the Mr. Frosty isopropanol-containing system.

CELL LINES

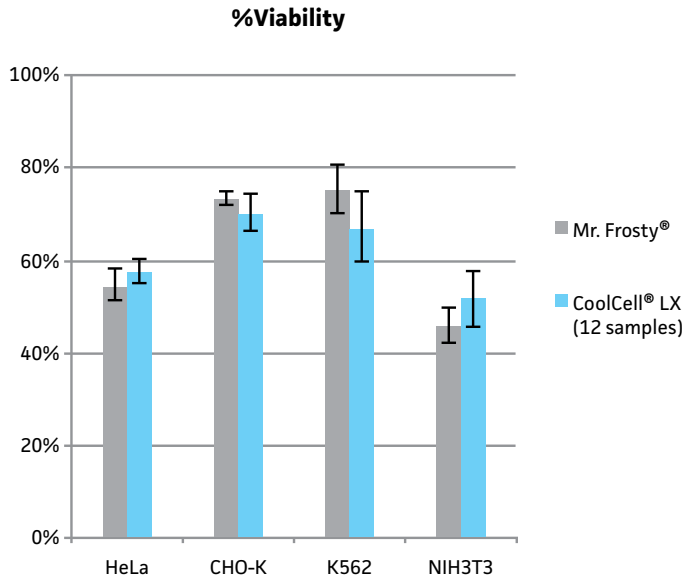


Figure 1. HeLA, CHO-K, K562, NIH3T3 were cryopreserved using the CoolCell® LX, or Mr. Frosty® freezing containers. All four cell lines performed equally well when post-thaw samples, from either container, were tested for viability and transfection efficiency.

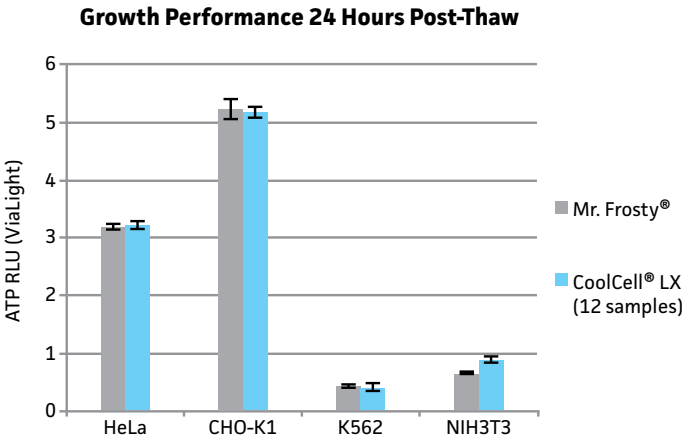


Figure 2. Post-thaw growth performance was similar for all of the cell lines tested, cryopreserved in either container.

PHONE
800.638.6597
703.365.2700

EMAIL
sales@atcc.org
tech@atcc.org

WEB
www.atcc.org

ATCC®

10801 University Blvd.
Manassas, VA 20110

CB-0513-02